

Discovery of a thermotolerant MHET hydrolase scaffold using bioinformatics and machine learning

K. Grigorakis¹, E. Nikolaiivits¹, C. Ferousi¹, E. Topakas^{1*}

¹Industrial Biotechnology & Biocatalysis Group, Biotechnology Laboratory, School of Chemical Engineering, National Technical University of Athens, Athens, Greece

(*vtopakas@chemeng.ntua.gr)

ABSTRACT

Polyethylene terephthalate (PET) is the most recycled plastic worldwide, the most widely used material for beverage packaging, and valuable enough to solicit R&D projects to its after-use recovery^[1]. Enzymatic depolymerization of PET to its monomeric units, i.e., terephthalic acid (TPA) and ethylene glycol (EG), allows for its separation from plastic mixtures, its valorization to valuable chemicals and the production of high-quality repolymerized PET, in contrast to material-degrading thermomechanical recycling methods^[2, 3]. Several reported PET hydrolases (PETases) enable this waste–PET biorefinery, but suffer inhibitory effects due to the formation of monohydroxyethyl terephthalate (MHET); an intermediate hydrolysis product^[4]. This inhibition can be rectified by introducing an MHET hydrolase enzyme (MHETase) that degrades MHET to TPA and EG^[5]. However, the optimal temperature of enzymatic PET degradation is close to its glass transition temperature of 70 °C^[6, 7], and while PETases and other esterases that can withstand these temperatures have been discovered and engineered before^[8, 9], no reported MHETases can withstand temperatures higher than 50 °C^[10]. In this work, putative thermotolerant MHETases are mined with the bioinformatics tools BLAST and machine learning-based ThermoProt^[11, 12]. Selection is based on similarity to the well-studied *IsMHETase* from *Ideonella sakaiensis*, predicted protein secretion type and thermophilicity. The most promising candidate is *ZcMHETase* from *Zhizhongheella caldifontis* that is recombinantly expressed in *E. coli* Shuffle T7 and purified to homogeneity. Its temperature and pH stabilities and optima are experimentally determined on pNP–butyrate substrate, and its MHET hydrolysis capacity is compared to *IsMHETase*. Protein engineering of *ZcMHETase* aiming at enhancing catalytic activity is currently ongoing, based on previously identified *IsMHETase* hot-spot residues and inspection of relevant crystal structures^[13]. Ultimately, *ZcMHETase* could be used as a new scaffold for designing more efficient enzymatic PET valorization and recycling systems.

KEYWORDS: MHETase, Thermotolerant, ThermoProt, Machine Learning, Bioinformatics

REFERENCES

- [1] Sarda P, Hanan JC, Lawrence JG, Allahkarami M (2022) *Journal of Polymer Science* 60:7–31
- [2] Kim HT, Kim JK, Cha HG, et al (2019) *ACS Sustain Chem Eng* 7:19396–19406
- [3] Kaabel S, Daniel Therien JP, Deschênes CE, Duncan D, Friščić T, Auclair K (2021) *Proc Natl Acad Sci U S A* 118:e2026452118
- [4] Barth M, Oeser T, Wei R, Then J, Schmidt J, Zimmermann W (2015) *Biochem Eng J* 93:222–228
- [5] Yoshida S, Hiraga K, Takehana T, Taniguchi I, Yamaji H, Maeda Y, Toyohara K, Miyamoto K, Kimura Y, Oda K (2016) *Science (1979)* 351:1196–1199
- [6] Ellis LD, Rorrer NA, Sullivan KP, Otto M, McGeehan JE, Román-Leshkov Y, Wierckx N, Beckham GT (2021) *Nature Catalysis* 2021 4:7 4:539–556
- [7] Wilkes CE, Summers JW, Daniels CA, Berard MT (2005) Hanser
- [8] Tournier V, Topham CM, Gilles A, et al (2020) *Nature* 2020 580:7802 580:216–219
- [9] Sun J, Pang Y, Lei Z, OuYang B, Lai W, Wang Y, Lan D (2024) *Biochem Eng J* 204:109222

- [10] Palm GJ, Reisky L, Böttcher D, Müller H, Michels EAP, Walczak MC, Berndt L, Weiss MS, Bornscheuer UT, Weber G (2019) *Nat Commun* 10:1717
- [11] Bateman A, Martin MJ, Orchard S, et al (2023) *Nucleic Acids Res* 51:D523–D531
- [12] Erickson E, Gado JE, Avilán L, et al (2022) *Nature Communications* 2022 13:1 13:1–15
- [13] Knott BC, Erickson E, Allen MD, et al (2020) *Proc Natl Acad Sci U S A* 117:25476–25485